



## Research Articles

# Comparative Histomorphologic Evaluation of the Testicular Apparatus of Vertebrates

**Odokuma Emmanuel I<sup>1</sup> and Ibeabuchi Nwachukwu M<sup>2</sup>**

<sup>1</sup>Department of Human Anatomy and Cell Biology, Delta State University, Abraka, Nigeria

<sup>2</sup>Department of Anatomy, College of Medicine, University of Lagos, Lagos, Nigeria

*\*Author for Correspondence:*

**Odokuma E. I.**

E-mail: secretfiles1800@yahoo.com

### Keywords:

Vertebrates,  
Amniotes,  
Phylogeny,  
Histology,  
Comparative

### SUMMARY

**Objectives:** The testis, the reproductive organ of male vertebrates, shares common features across the Amniotes, Phylum. While variations are well circumscribed, only sparse documentation on comparative histomorphology of the vertebrate testes exists and these are focused mainly on within-class interspecies Phylogeny characterization.

**Methods:** Ethical clearance for this comparative histologic observational study was obtained from the HREC of the Department of Human Anatomy and Cell Biology, Delta State University, Abraka (No. DELSU/CHS/ANA/118). The research was conducted in accordance with the guidelines for the care and use of animals for research.

**Results:** The testes of the studied vertebrates all possessed capsules which was thin in non-amniotes and thicker in amniotes. Seminiferous tubules were common to all classes though cystic patterns were peculiar to fish and amphibians. It was revealed that fish had the least developed tubular system and number of germ cells per tubule while germ cells were most abundant in birds. Sertoli cells were present in all the seminiferous tubules of the vertebrate classes.

**Conclusion:** The reported histomorphologic features in the studied vertebrates strongly demonstrate testicular characteristics that are conserved within the group despite differences in class habitat. The implication is the probable existence of a tightly regulated genetic control of male gonads within the phylum. The findings suggest that the observed differences arose from possible adaptational changes owing to evolutionary pressures within the different environments to which the animal class found itself.

## INTRODUCTION

The testes are the reproductive organ of male vertebrates, displaying conserved features across the phylum, albeit apparent variations.[1] The germ cells have earlier been shown to originate from the wall of the yolk sac in early embryonic life and migrate to the final position as observed in the adult.[2] The testes not only accommodates the germ cells but is also the location for Sustentacular cells of Sertoli and interstitial cells of Leydig which are instrumental to development of male phenotypic characteristics through its production of the hormone, testosterone, the most potent natural androgen.[3] The male sex cells, the spermatozoa are generated in the seminiferous tubules of the testes through empirical processes of mitosis and meiosis, from their precursors; spermatogonia, spermatocytes and spermatids.[4]

The presence of a vertebral column derived from a notochord is the anatomical basis for taxonomical designation of the vertebrate phylum.[5] Despite this long accepted characteristic, vertebrates share other features peculiar to the group. A copious literature search scarcely revealed the effects of adaptational and/or evolutionary factors in relatedness vis-à-vis variations existing between and within vertebrates.[5-10]

The thrust of this study therefore was to demonstrate the features of the testes in vertebrates and provide a detailed histologic description of the testes of the different classes. The observed ontogenic patterns will definitely provide an explanation for the relatedness of members of the vertebrate phylum. The possible evolutionary, adaptational factors involved and the histomorphologic variations existing in the vertebrate phylum was also established.

## METHODS

Permission for this comparative histologic observational study was sought and received from the research and ethics committee of the department of Human Anatomy and Cell Biology, Faculty of Basic Medical Sciences, College of Health Sciences, Delta State University, Abraka (Number DELSU/CHS/ANA/118). The research was conducted in accordance with the guidelines for the care and use of animals for research.[11-12]

Five animals were utilised for the study (one from each vertebrate class). The animals included: common cat fish (*Clarias gariepinus* - fish); the toad (*Bufo bufo* - amphibian); the agama lizard (*Agama agama* - reptile); the house hold chicken (*Gallus domesticus* - bird) and the Wistar rat (*Rattus norvegicus* - mammal).

Following euthanasia, the testes of each animal were dissected out and preserved in 10% formal saline for about 48 hours. The samples were then processed using standard procedures involving progressive dehydration from 75% to absolute alcohol, emersion in xylene, embedding in wax, processing through sectioning (3-4µm) and staining with haematoxylin and eosin.[13] The histology slides obtained were then analysed and interpreted with the aid of a Scopetek DCM 500, 5.0 mega pixel Microscope Camera and computer.

## RESULTS

Sections of the testes of a fish show a capsule (Figures 1), 1-2 layer thick of dense connective tissue in which are some duct-like structures lined by pseudostratified ciliated epithelium (Figure 2). The capsule encloses several diverse shaped cystic structures lined by moderately sized germ cells with round to oval centrally disposed bland nuclei and pale cytoplasmic borders (Figure 3). These stem cells give rise to about four to five generation of cells with smaller size as they approach the luminal aspect of the cyst (Table 1). The cells are held together by extensive cytoplasmic processes of Sertoli cells which possess a small round to oval nuclei, a pattern suggestive of a monoclonal origin. Each cystic structure is separated by a fibrovascular connective tissue stroma in which are few pale interstitial cells with vesicular nuclei disposed in clusters. These cells are relatively large with pale nuclei.

Sections of toad testes showing several variably sized cystic structures lined by flattened epithelial cells with bland spindle shaped nuclei (Figure 4). The cysts invest germs cells of varying stages of maturity, disposed in clusters invested by cytoplasmic processes of Sertoli cells which possess round to oval pale nuclei with prominent nucleoli (Figure 5). Corresponding with Sertoli cells, about 4-6 clusters of cells are present in each cyst 4-6 layers thick (Table 1). The terminally differentiated cells are flagellated and invested in cytoplasmic processes forming pseudo-rosettes. Pale cells are intermixed in the intervening fibrovascular connective tissue stroma (Figure 6). These Leydig cells possess vesicular variably shaped nuclei. There is a 1 to 2 cell layer of fibroconnective tissue stroma investing the testes.

Sections showing several seminiferous tubules of varying sizes in a lizard testis (figure 7). Each tubule is composed of germinal epithelium at different stages of development (figure 8). The basal spermatogonia are

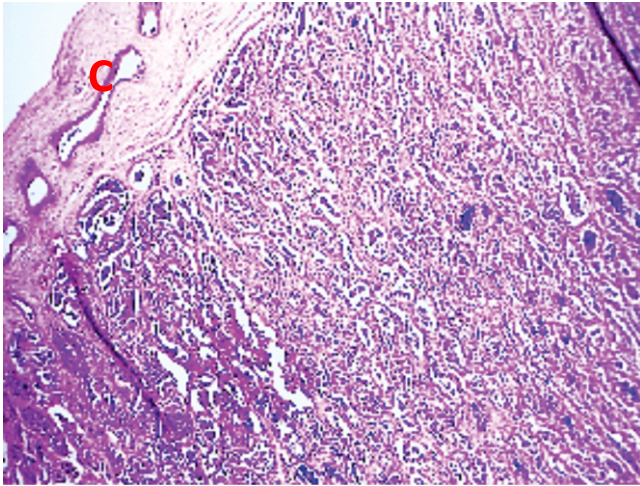
relatively small with dense nuclei and scant cytoplasm. This gives rise to the slightly larger primary spermatocytes, spermatids and flagellated spermatozoa which are in the luminal aspect of tubule. The cells occur in columns with about 15-20 columns, corresponding with Sertoli cells, per tubule and 10-15 cells per column (Figure 9, Table 1). The intervening fibrovascular stroma contains moderately sized Leydig cells with squamoid appearance and vesicular nuclei. There is a 2-3 cell layer dense fibroconnective tissue capsule investing the testes.

Sections of bird testes show several variably sized seminiferous tubules disposed within a fibrovascular stroma in which are several moderately sized interstitial cells (Figure 10). These Leydig cells possess pale cytoplasm with moderate sized nuclei and prominent nucleoli. Within the tubule are germ cells at different stages of development (Figure 11). The cells are arranged in columns, coinciding with the support cells (Sertoli cells) with each tubule containing 20-25 of such columns and each column consisting of 5-10 cells (Table 1). The basal cells, spermatogonia display nucleopleomorphism with mild vesiculation and scant cytoplasm. 7-8 generations of cells have eosinophilic cytoplasm with evidence of mitotic changes occurring (Figure 12). The spermatids, small cells with small sized nuclei and eosinophilic cytoplasm are present with flagella tails disposed in the luminal aspect of the tubule.

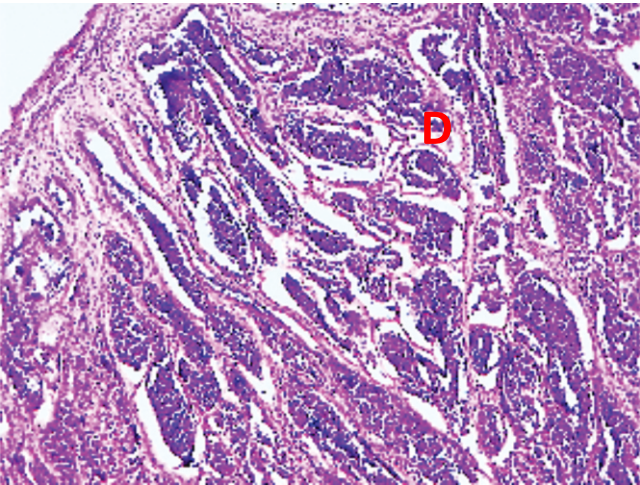
Sections of rat testes indicate several variably sized seminiferous tubules lined by germ cells at different stages of maturation (Figure 13). Spermatogonia, small sized cells with oblong, densely coloured nuclei line the basal aspect of each seminiferous tubule (Figure 14). Primary spermatocytes follow these stem cells and they are committed to production of spermatids. The primary spermatocytes are moderately large cells with moderately abundant eosinophilic nucleus and with chromatin in mitosis. Spermatids are relatively smaller flagellated cells held together by cytoplasmic processes. From the spermatogonia to the terminally differentiated spermatozoa, the cell layer is about 6-10 layer thick in most tubules (Figure 15, Table 1). Interstitial cells of Leydig are present in the intervening fibromuscular connective tissue stroma. Blood vessels are also seen. The testis is invested in a dense regular 2-3 cell layer thick connective tissue capsule.

**Table 1: Vertebrate testes; Comparison of Microanatomic Structure**

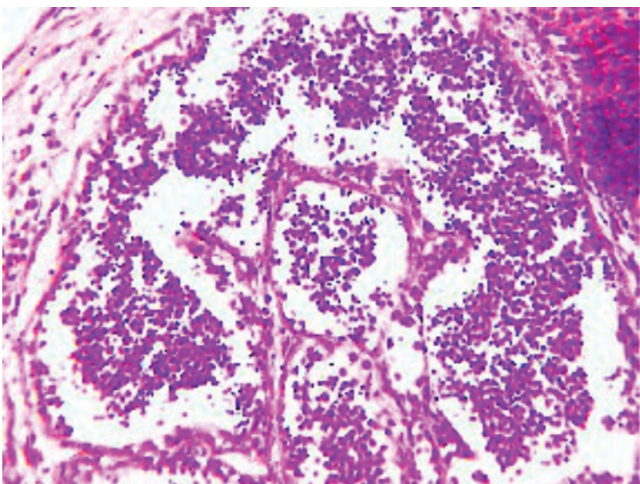
| S/N | Vertebrate | Capsule  | Interstitial | Seminiferous Tubule Pattern | Cell location | Cell arrangement | Flagella - |
|-----|------------|----------|--------------|-----------------------------|---------------|------------------|------------|
| 1.  | Fish       | Thin     | Leydig cells | Cystic                      | Central       | Singles/clusters | +          |
| 2.  | Amphibian  | Thin     | Leydig cells | Cystic                      | Peripheral    | Clusters 4-6     | +          |
| 3.  | Reptile    | Moderate | Leydig cells | Small tubules               | Peripheral    | Columns 10-15    | +          |
| 4.  | Bird       | Thick    | Leydig cells | Polygonal tubules           | Peripheral    | Columns 5-10     | +          |
| 5.  | Mammal     | Thick    | Leydig cells | Round to Oval tubules       | Peripheral    | Columns 6-10     | +          |



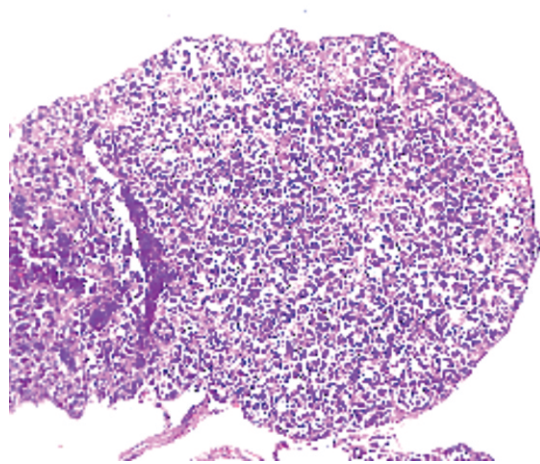
**Figure 1:** Photomicrographic section of the testes of cat-fish *Clarias gariepinus* showing the Capsule C. (H&E stain, mag x40).



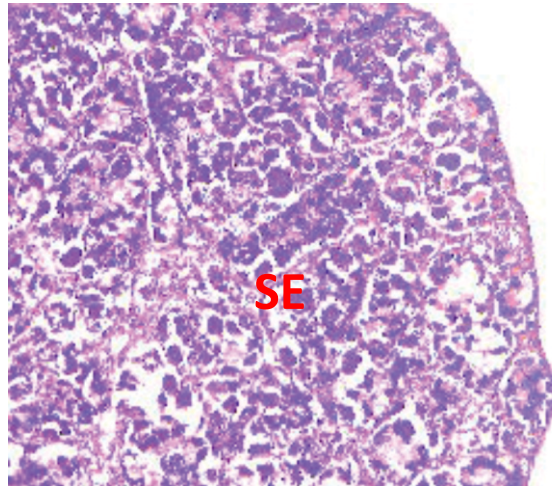
**Figure 2:** Photomicrographic section of the testes of cat-fish *Clarias gariepinus* showing duct-like structures, D, lined by pseudostratified ciliated epithelium (H&E stain, mag x100).



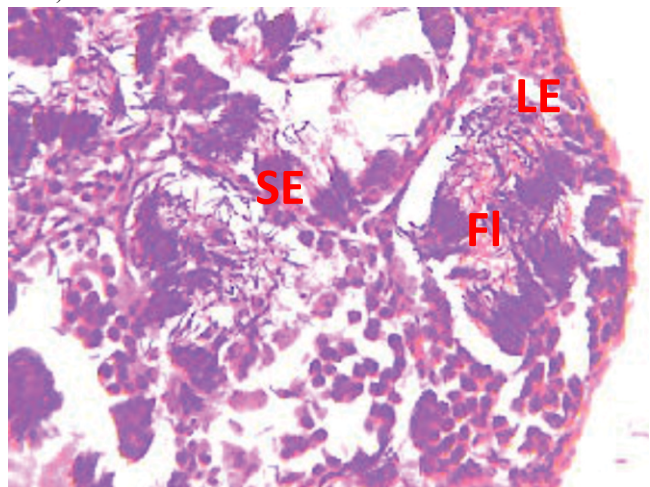
**Figure 3:** Photomicrographic section of the testes of cat-fish *Clarias gariepinus* showing the capsule enclosing several diverse shaped cystic structures, Cy, and lined by moderately sized germ cells with round to oval centrally- disposed bland nuclei and pale cytoplasmic borders (H&E stain, mag x400).



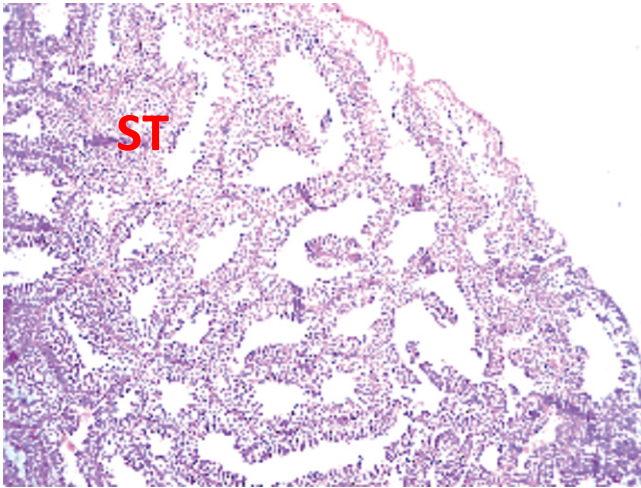
**Figure 4:** Photomicrographic section of the testes of the amphibian toad *Bufobufo* showing several variably sized cystic structures lined by flattened epithelial cells with bland spindle shaped nuclei (H&E stain, mag x40).



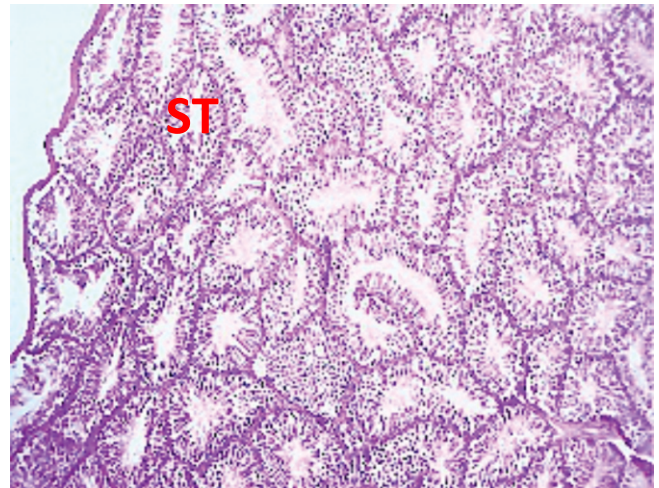
**Figure 5:** Photomicrographic section of the testes of amphibian toad *Bufobufo*. The cysts, Cy, invest germs cells of varying stages of maturity, disposed in clusters invested by cytoplasmic processes of Sertoli cells, SE. Corresponding with Sertoli cells, about 4-6 clusters of cells are present in each cyst 4-6 layers thick (H&E stain, mag x100).



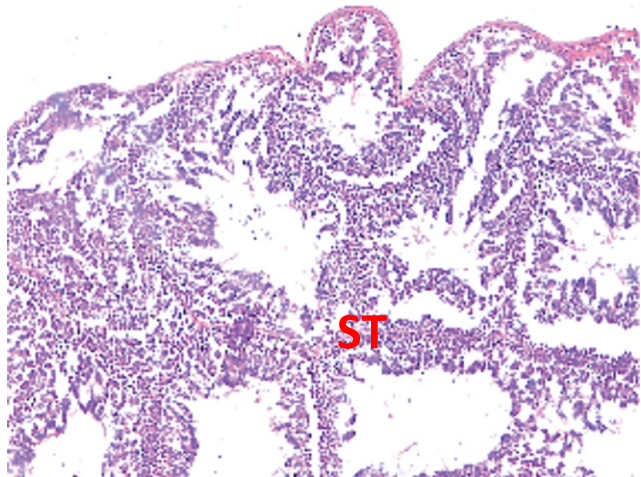
**Figure 6:** Photomicrographic section of the testes of amphibian toad *Bufobufo*. Pale cells, LE, are intermixed in the intervening fibrovascular connective tissue stroma. Flagellated cells, FI, are also seen (H&E stain, mag x400).



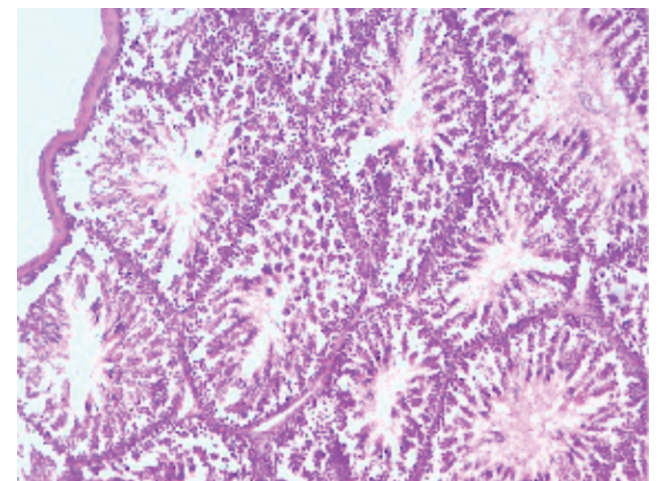
**Figure 7:** Photomicrographic section showing several seminiferous tubules, ST, of varying sizes in a reptilian lizard *Agama agama*, testis (H&E stain, mag x40).



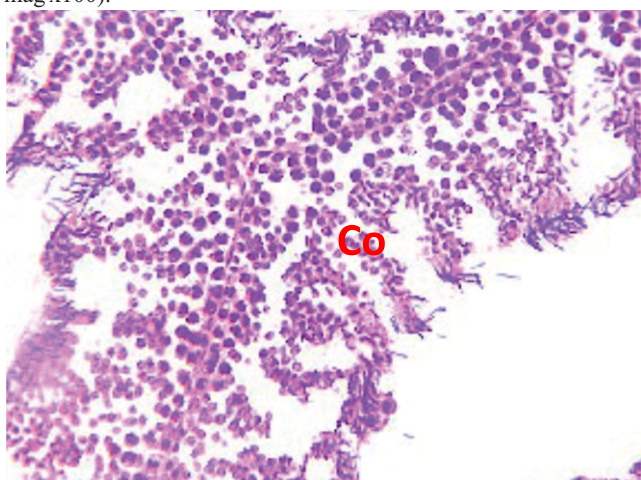
**Figure 10:** Photomicrographic section of bird, domestic fowl *Gallus domesticus*, testes showing several variably sized seminiferous tubules disposed within a fibrovascular stroma in which are several moderately sized interstitial cells (H&E stain, mag x40).



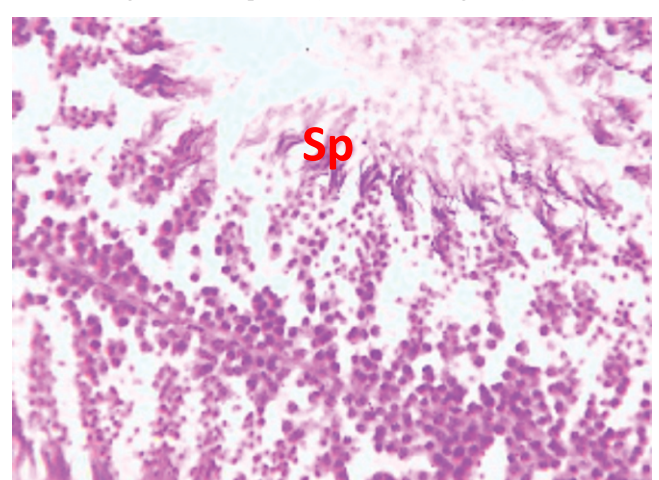
**Figure 8:** Photomicrographic section of the testes of reptilian lizard *Agama agama*. Each tubule is composed of germinal epithelia at different stages of development. The basal spermatogonia are relatively small with dense nuclei and scant cytoplasm (H&E stain, mag x100).



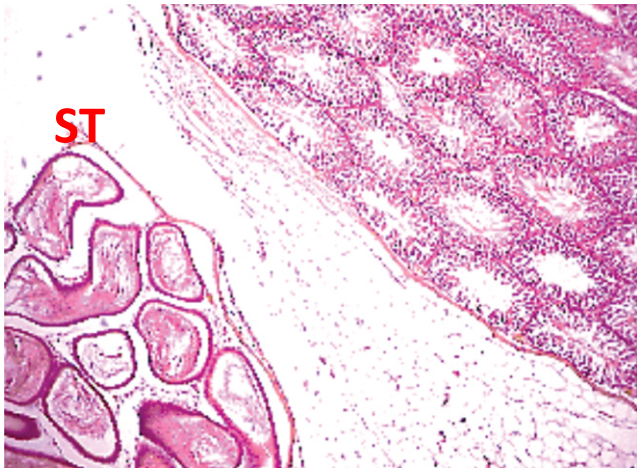
**Figure 11:** Photomicrographic section of the testes of bird, domestic fowl *Gallus domesticus*. Within the tubules are germ cells at different stages of development (H&E stain, mag x100).



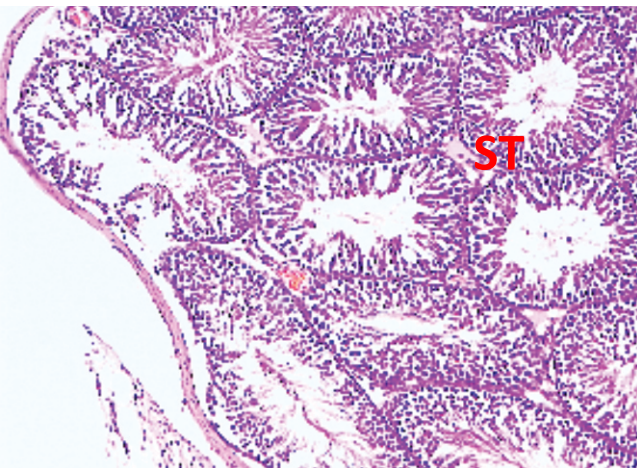
**Figure 9:** Photomicrographic section of the testes of reptilian lizard *Agama agama*. The intervening fibrovascular stroma contains moderately sized Leydig cells with squamoid appearance and vesicular nuclei. There is a 2-3 cell layer dense fibroconnective tissue capsule investing the testes (H&E stain, mag x400).



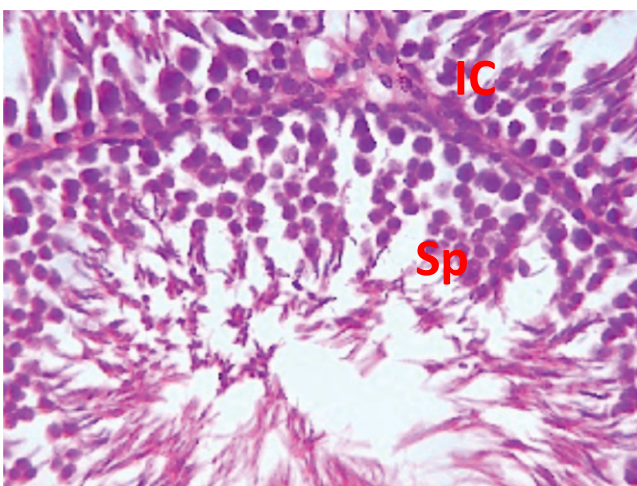
**Figure 12:** Photomicrographic section of the testes of bird, domestic fowl *Gallus domesticus*. The cells are arranged in columns, coinciding with the support cells (Sertoli cells) with each tubule containing 20-25 of such columns and each column consisting of 5-10 cells. The spermatids, small cells with small sized nuclei and eosinophilic cytoplasm are present with flagella tails disposed in the luminal aspect of the tubule (H&E stain, mag x400).



**Figure 13:** Photomicrographic section of mammalian Wistar rat, *Rattus norvegicus* testes indicating several variably sized seminiferous tubules, ST, lined by germ cells at different stages of maturation. At the lower left corner are sections of epididymis (H&E stain, mag x40).



**Figure 14:** Photomicrographic section of the testes of mammalian Wistar rat, *Rattus norvegicus*. Spermatogonia, small sized cells with oblong, densely coloured nuclei line the basal aspect of each seminiferous tubule. Spermatids are relatively smaller flagellated cells held together by cytoplasmic processes. (H&E stain, mag x100)



**Figure 15:** Photomicrographic section of the testes of mammalian Wistar rat, *Rattus norvegicus*. From the spermatogonia to the terminally differentiated spermatozoa, Sp, the cell layer is about 6-10 layer thick in most tubules. Interstitial cells, IC, of Leydig are present in the intervening fibromuscular connective tissue stroma. Blood vessels are also seen. The testis is invested in a dense regular 2-3 cell layer thick connective tissue capsule (H&E stain, mag x400).

## DISCUSSION

This study has elucidated the structural relationships and variations existing between and within the male reproductive testes in vertebrates. The observations are strongly suggestive of interplay between both adaptive and evolutionary factors in fashioning of ontogenic patterns of testicular morphology in the studied vertebrate subphylum. Assessment of fish testes revealed that they possessed the most primitive form of this male sex organ in the group. The lobulocystic patterns observed in this study was not new. Several authorities have affirmed that some fish species displayed cystic lobular arrangement - a feature of primitive gonads.[7,14-17] It has been further suggested that some fish species, teleosts, maintained testicular cycles in which two sets of tubules were present in the same fish, some lobules containing spermatogenic cells while others were clustered with differentiated/specialised spermatozoa.[16,18-20]

Only spermatogenic cells, spermatogonia and spermatocytes were described in this study. It is likely that the fish analysed was at the period of the testicular cycle identified by Ahmed *et al.* as spermatogenic testis in which most of the luminal cells were in the developing mitotic phase of the cell cycle unlike the pubertal testis in which the testes was in the meiotic phase.[7,21] Another important observation was the clustering of cells within each cyst. Cell clusters were invested in cytoplasmic processes of Sertoli cells which have been shown to provide nutrition for the spermatogenic cells.[2]

A thin fibroconnective tissue capsule was noted encasing the fish testes. The importance of this is not farfetched especially as the mechanism of release of the germ cells from the male sex organ is by rupturing of the cystic tubules into the surrounding fluid medium and migration of the spermatozoa to the eggs hitherto laid by the female fish. The toad, amphibian was also observed to possess a similar thin fibroconnective tissue capsule in which the testes were invested. Some authors have confirmed that most amphibians possessed extensive seminiferous tubules in their testes, each containing clusters of clonal germ cells whose nutrition is provided by a Sertoli cell. [22-23] As was demonstrated in the index study, most clones were composed of columns of between 4-6 cells of the spermatogenic series like Rastogi *et al.* and Takamure *et al.* had previously confirmed.[22-25]

The existence of monoclonal progenies of spermatogonia in each tubule had earlier been identified [26-28] and the current study further demonstrated an extensively populated seminiferous tubule, with more flagellated spermatogonia in the toad than in the fish. Though both the fish and toad possessed cystic structures for generating germ cells, several factors had the tendency of affecting amphibians more. Rastogi *et al.*, and Loumbourdis and Kyrie corroborated this fact when they affirmed the influence of environmental fluctuations on the testes of toads.[29,30] The fish on the other hand inhabit a more stable aquatic environment which is indispensable in the dispersal of spermatozoa for effective fertilization.

The interstitial cells described in this study were not new. A prior investigation by Fraile *et al.*[31] had shown a parenchymal to stromal ratio of 7:1 which was in keeping with the finding in this study even amongst vertebrates of higher classes. This apparent conservation of interstitial/stromal tissue also extended specifically to the

Leydig cell morphology which also clearly exhibited close similarity in the vertebrate phylum.

Reptiles, unlike fish and amphibians, are the first in the series of amniotes. This group of vertebrates are adapted to the terrestrial environment by developing seminiferous tubules with numerous germ cells and further developed Sertoli cells which are retained following release of the mature germ cells from the tubules.[6,32-34]

Generally, the structure of the seminiferous tubule could clearly be differentiated from those of fish and amphibians. They were larger and exhibited a close association between each germ cell cluster and Sertoli cell like in mammals and birds. Russel *et al.* had previously emphasised this spatial arrangement.[35] The significance of this pattern has been ascribed to internal fertilization, an evolutionary sophistication of amniotes.[36-37] Despite the inclination to the mammalian organisation, the cluster/tubular inconsistency of the amphibians were however retained.[38-44] The interstitial cells were not different from those of the fish and amphibians further corroborating an earlier reports that the Leydig cell morphology was conserved throughout the vertebrate phylum.[45-46]

The seminiferous tubules of birds were distinctly polypoid in shape with prominent interstitial cells of Leydig. The observed abundant Leydig interstitial cells observed could have been a result of seasonal changes in the reproductive cycle of the bird studied. Rosenstrauch *et al.* [47] revealed that interstitial cells were more prominent during breeding than in non-breeding states. The specific features of these cells, prominent nucleoli within a vesicular nucleus in a squamoid cell with granulated cytoplasm, as seen in other lower vertebrates, remained unaltered.

Germ cells at different stages of maturation lined the tubules. Each column was closely associated with its own sustentacular cell. Paramount was the multiple stages/generation of differentiation of the germ cells unlike in the reptiles, Amphibians or fish. Also, several of such columns per tubule were observed. Pudney[48] opined that this was necessary for adequate reservoir for subsequent spermatogenic activity following ejaculation.

Another striking feature of the testes in the bird studied was the dense fibroconnective tissue investing the male gonad. This dense irregular connective tissue was observed to extend deep between and within the seminiferous tubules, separating them into clusters, albeit less intense than the thickness of the overlying capsule. Aire and Ozegbe [49] explained that myoid tissue present in bird capsule was of significant importance in aiding the propulsion of fluid content of the testes to the epididymis. Cortical and medullary septa from the capsule were however not present.[50-52]

Similarly, the testes of the mammal,(rat), was observed to be enclosed by a similar dense irregular connective tissue capsule which displayed extensions into the testicular parenchyma carrying neurovascular bundles providing nourishment. Unlike the polygonal disposition of the bird seminiferous tubules, those of rats were round to oval in shape and surrounded by interstitial cells of Leydig. These cells, as has earlier been described were not different from those of the other vertebrates in the phylum.

In conclusion, the reported histomorphologic features

in the studied vertebrates have strongly suggested characteristics that are conserved within the group despite differences in class habitat. The implication of this is that there exists a genetic control of male gonads within the phylum which is tightly regulated. Also, the existence of differences, which has most likely developed from adaptational changes owing to the different environments to which the animal class finds itself, has been established.

## ACKNOWLEDGEMENT

The authors are most indebted to Agbabi CO, who assisted with data collection and Chukura UC, who was very helpful with the editorial work, in this publication.

## Declaration

We declare no conflict of interest.

## REFERENCES

1. Defalco T, Capel B. Gonad Morphogenesis in Vertebrates: Divergent Means to a Convergent End. *Annu Rev Cell Biol.* 2009;25:457-482.
2. Sadler TW Langman's Medical Embryology 8th ed Lippincott William and Wilkins Philadelphia, Pennsylvania. 2000;Ch14:Pp 321.
3. Karl J, Capel B. Sertoli cells of the mouse testis originate from the coelomic epithelium. *Dev Biol.* 1998;203:323-333.
4. Sadler TW Langman's Medical Embryology 8th ed Lippincott William and Wilkins Philadelphia, Pennsylvania. 2000;Ch14 pp4-8.
5. Fleming A, Keynes RJ, Tannahill D The role of the notochord in vertebral column formation *J. Anat.* 2001;199:177-180.
6. Gribbins KM. Reptilian Spermatogenesis; A histological and Ultrastructural Perspective. *Spermatogenesis.* 2011;1(3):250-269.
7. Ahmed YA, Abdel Samei NA, Zayed AZ Morphological and Histomorphological Structure of Testes of the Catfish '*Clarius gariepinus*' from Egypt. *Pak J Biol Sci.* 2013;16(13):624-629.
8. Meisner AD, Burns JR, Weitzman SH, Malabarba LR. Morphology and Histology of the male reproductive system in two species of internally inseminating South American Catfishes *Trachelyopterus lucenai* and *T. galeatus*. *J Morphol.* 2000;246:313-342
9. Roosen-Runge EC. The Process of Spermatogenesis in Animals. Cambridge, England: Cambridge University Press; 1977. pp. 5-250.
10. Senger K. Transcriptional and translational regulation of gene expression in haploid spermatids. *Anat. Embryol.* 1999;199:471-487.
11. Helsinki R, Thomas K. Research and Research Ethics: 2002;1-4.
12. Yorezawa M, Yamazaki H, Shimada T. Animal Model for Research: 2010; pp70-72.
13. Clark G. Staining procedures. *Instructional Guide* 2003;4:412-419.
14. Chaves-Pozo E, Mulero V, Meseguer J, Ayala AG. An Overview of Cell renewal in the testis throughout the reproductive cycle of a seasonal breeding teleost, the gilthead seabream (*Sparus aurata* L). *Biol Reprod.*

- 2005;72:593-601.
15. Loir M, Cauty C, Planquette P, Le Bail PY. Comparative study of the male reproductive tract in seven families of South-American catfishes. *Journal of Aquatic Living Resources*. 1980;2:45-56.
16. Suwanjarat J, Amornsakun T, Thongboon L, Boonyoung P. Seasonal changes of spermatogenesis in the male sand goby *Oxyeleotris marmoratus* Bleeker, 1852 (Teleostei, Gobiidae). *Songklanakarin J Sci Technol*. 2005;27:425-436.
17. VanDyk JC, Pieterse GM. A histo-morphological study of the testis of the sharp tooth catfish (*Clarias gariepinus*) as reference for future toxicological assessment. *J Applied Ichthyol*. 2008;24:415-422.
18. Cek S, Yilmaz E. gonad development and sex ratio of sharp tooth catfish *Clarias gariepinus* (Burchell,1822). Cultured under Laboratory Conditions. *Turk J Zool*. 2007;1:35-46.
19. Lawson EO. Testicular maturation and reproductive cycle in mudskipper, *Periophthalmus papillo* (Blochand Schneider 1801) from Lagos Lagoon, Nigeria. *J of Am Sci*. 2011;7: 4859.
20. Shrestha TK, Khanna SS. Seasonal changes in the testes of a hill stream teleost, *Garragotyla* (Gray) *Acta Anat (Basel)*. 1978;100:210-220.
21. Emam MA, Abughrien B. Seasonal Histology Changes in Gonads of the Catfish. *Fish Aquac J*. 2014;5(1):1-5.
22. Trauth SE, Thigpen CS, Conner MB Testicular Histology and sperm Morphometrics of the Bird-Voiced Treefrog, *Hylaavivoca* (Anura:Hylidae), from Arkansas. *J Arkansas Acad of Sci*. 2015;69:101-105.
23. Dodd CK Jr. *Frogs of the United States and Canada*. Vols. 1 and 2. The Johns Hopkins University Press, Baltimore (MD). 2013; pp982.
24. Rastogi RK, Bagnara JT, Iela L, Krasovich MA. Reproduction in the Mexican leaf frog, *Pachymedusa dacnicolor*. IV. Spermatogenesis: a light and ultrasonic study. *J Morph*. 1988;197:277-302.
25. Takamune K, Teshima K, Maeda M and Abé S. Characteristic features of preleptotene spermatocytes in *Xenopus laevis*: increase in the nuclear volume and first appearance of flattened vesicles in these cells. *The J Exp Zool*. 1995;273:264-270.
26. Chavadej J, Jerareungrattana A, Sretarugsa P, Sobhon P. Structure and Development of the Testis of Bullfrog, *Rana catesbeiana*, and Their Changes during Seasonal Variation. *Science Asia*. 2000;26:69-80.
27. Kalt MR Morphology and kinetics of spermatogenesis in *Xenopus laevis*. *J Exp Zool*. 1976;195:393-408.
28. Callard IP, Callard GV, Lance V, Bolaffi JL and Rosset JS. Testicular regulation in non-mammalian vertebrates. *Biol Reprod*. 1978;18:16-43
29. Rastogi RK, Iela L, Delrio G, Dimeglio M, Russo A and Chieffi G. Environmental influence on testicular activity in the green frog *R. esculenta*. *J Exp Zool*. 1978;206:49-64.
30. Loumbourdis NS and Kyriakopoulou-Sklavounou P. Reproductive and lipid cycles in the male frog *Rana ridibunda* in northern Greece. *Comp Biochem Physiol*. 1991;99: 557-83.
31. Fraile F, Paniagua R, Saez FJ, Rodriguez MC, Lizana M. Immunocytochemical and quantitative study of interstitial cells in the high mountain toad *Bufo bufo gredosicola* during the spermatogenic cycle. *Herp J*. 1989;1:330-335.
32. Bergmann M, Schindelmeiser J, Greven H. The blood-testis barrier in vertebrates having different testicular organizations. *Cell Tiss Res*. 1984;238:145-150.
33. Baccetti B, Bigliardi E, Talluri MV, Burrini AG. The Sertoli Cell in lizards. *J Ultrastruct Res*. 1983;168:268-275.
34. Gribbins KM. The cytological evaluation of spermatogenesis and ultrastructure of the junctional complexes within the germinal epithelium of reptiles. Cincinnati OH: University of Cincinnati; 2003. Doctoral Thesis.
35. Russell LD, Hikim SAP, Ettlin RA, Clegg ED. *Histological and Histopathological evaluation of the testis*. Clearwater FL: Cache River Press; 1990.
36. Pudney J. Comparative cytology of the non-mammalian vertebrate Sertoli cell. In: Russell LD, Griswold MD, editors. *The Sertoli Cell*. Clearwater FL: Cache River Press; 1990. pp.611-657.
37. Gist DH, Turner TW, Congdon JD. Chemical and thermal effects on the motility of sperm from the turtle epididymis. *J Reprod Fert*. 2000;119:271-277.
38. Gribbins KM, Gist DH, Congdon JH. Cytological evaluation of spermatogenesis and organization of the germinal epithelium in the male Slider Turtle, *Trachemys scripta*. *J Morphol*. 2003;255:337-346.
39. Gribbins KM, Elsey R, Gist DH. Cytological evaluation of spermatogenesis in the germinal epithelium of the American Alligator, *Alligator mississippiensis*. *Acta Zool*. 2006;87:59-69.
40. Gribbins KM, Gist DH. The cytological evaluation of the germinal epithelium and the germ cell cycle in an introduced population of European Wall Lizard, *Podarcis muralis*. *J Morphol*. 2003;256:296-306.
41. Gribbins KM, Happ CS, Sever DM. Ultrastructure of the reproductive system of the Black Swamp Snake (*Seminatrix pygaea*) V. The temporal germ cell development strategy of the testis. *Acta Zool*. 2005;86:223-230.
42. Gribbins K, Rheubert J, Collier M, Siegel D, Sever D. Histological analysis of spermatogenesis and the germ cell development strategy within the testis of the male western Cottonmouth Snake, *Agkistrodon piscivorus*. *Annal Anat*. 2008;190:461-476.
43. Gribbins KM, Rheubert JL, Poldemann EH, Collier MH, Wilson B, Wolf K. Continuous spermatogenesis and the germ cell development strategy within the testis of the Jamaican Gray Anole, *Anolis lineatopus*. *Theriogenology*. 2009;72:54-61.
44. Rheubert JL, McHugh HH, Collier MH, Sever DM, Gribbins KM. Temporal germ cell development strategy during spermatogenesis within the testis of the Ground Skink, *Scincella lateralis* (Sauria: Scincidae) *Theriogenology*. 2009;72:484-492.

45. Setchell BP. Male reproductive organs and semen. In: CUPPS, P. T. *Reproduction in Domestic Animals*, 4th edition. Academic Press, San Diego. 1991;pp.221-250.
46. Nora RI and Bertona MC. Seasonal changes in testicular activity of the protected *Boa occidentalis*, (Serpentes: Boidae): A histological study. *South Am J Herpetol.* 2006;43:143-148.
47. Rosenstrauch A, Weil S, Degen AA, Friedlander M. Leydig cell function, structure and plasma androgen level during the decline in fertility in aging roosters. *Gen Comp Endocrinol.* 1998;109(2):251-258.
48. Pudney J. Spermatogenesis in non-mammalian vertebrates. *Microscopy Research and Techniques.* 1995;32:459-497.
49. Aire TA, Ozegbe PC. The testicular capsule and peritubular tissue of birds: Morphometry, histology, ultrastructure and immunohistochemistry. *J Anat.* 2007;210:731-740.
50. Hodges RD. *The Histology of the Fowl*. London: Academic Press 1974
51. Lake PE. The male in reproduction. In: Bell DJ, Freeman BM, editors. *Physiology and Biochemistry of the Domestic Fowl*. New York: Academic Press. 1971;3:1411-1447.
52. Lake PE. Male genital organs. In: King AS, McLelland J, editors. *Form and Functions in Birds*. London: Academic Press. 1981;2:1-61.